

## Disjunct habitats as islands: genetic variability in the Caucasian rock lizard *Lacerta portschinskii*

Ross D. MacCulloch<sup>1</sup>, Robert W. Murphy<sup>1</sup>, Jinzhong Fu<sup>1</sup>, Ilya S. Darevsky<sup>2</sup> & Felix Danielyan<sup>3</sup>

<sup>1</sup>Centre for Biodiversity and Conservation Biology, Royal Ontario Museum, 100 Queen's Park, Toronto, Ontario M5S 2C6, Canada; <sup>2</sup>Zoological Institute, Russian Academy of Sciences, St. Petersburg 119034, Russia; <sup>3</sup>Faculty of Biology, Yerevan State University, 375000 Yerevan, Armenia

Received 28 October 1996 Accepted 1 April 1997

**Key words:** *Lacerta*, population genetics, protein electrophoresis

### Abstract

Genetic diversity at 37 allozyme loci was surveyed in *Lacerta portschinskii* from contiguous populations and from a disjunct population. Indices of genetic diversity (heterozygosity, number of alleles per locus, and percentage of loci polymorphic) were greater in contiguous populations than in the smaller disjunct population. In this regard, disjunct populations appear to be similar to island populations. Indices of genetic diversity in Caucasian *Lacerta* are less than those reported from vagile lizard taxa and more similar to those of sit-and-wait predators.

### Introduction

One factor affecting the amount of intrapopulation genetic variation is the size of the population. Reduction in genetic variability has been attributed to restriction of population size by geographic barriers and to founder effect. In one study, indices of genetic variability (heterozygosity and percentage of loci exhibiting polymorphism) in Adriatic island and mainland lacertid lizards (*Podarcis*) were found to be greater in mainland populations than in island populations (Gorman et al., 1975). Island size was also found to be positively correlated with variability, with indices of genetic variability of populations on larger islands approaching the values exhibited by mainland populations.

The greatest taxonomic diversity of the lizard genus *Lacerta* occurs in the Caucasian region (Darevsky, 1967). The varied topography and vegetation of the region have influenced the distributions of many species of *Lacerta*, producing some patchy, disjunct distributions.

An earlier study examined genetic variability in three species of Caucasian *Lacerta* (MacCulloch et al., 1995a). Species collected from restricted, disjunct

populations (*L. portschinskii*, *L. valentini*) exhibited lower values of genetic variability than did the species collected from a contiguous population (*L. rudis*). Although interspecific, the comparison was of some value because these three species are closely related, forming a well-supported clade in the phylogeny of *Lacerta* (Fu et al., in press; Murphy et al., 1996a).

Recently, *L. portschinskii* were collected in central Georgia, in the largest, contiguous portion of the species' range (Darevsky, Kupriyanova & Uzzell, 1985; MacCulloch et al., 1995a). This permitted a direct, intraspecific test of the hypothesis that geographically restricted disjunct populations are less variable genetically than are contiguous populations and thus resemble island populations, which are less variable than mainland populations.

### Materials and methods

Specimens were collected from three locations: Zedazeni, Manglisi, and Kodjori, within the largest, contiguous range of *Lacerta portschinskii* in central Georgia during 1995 (Figure 1). Specimens were

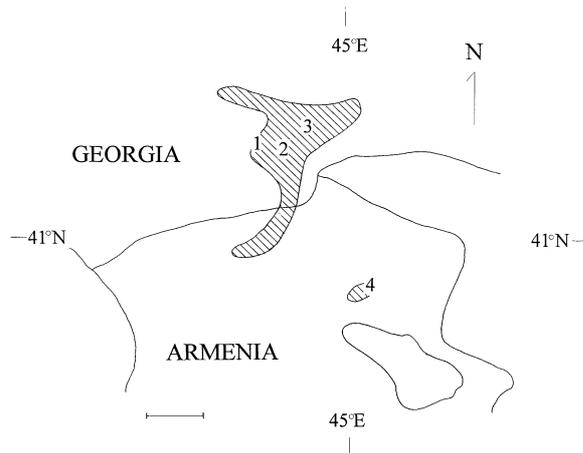


Figure 1. Distribution of *Lacerta portschinskii* (hatched area). Numbers represent collecting locations: (1) Manglisi; (2) Kodjori; (3) Zedazeni; (4) Gosh. The bar at the lower left represents 25 km.

euthanised by an overdose of sodium pentobarbitol and dissected immediately following euthanasia. Liver, heart, and skeletal muscle were removed and frozen in liquid nitrogen. Specimens are deposited in the herpetological collection of the Royal Ontario Museum (ROM; Appendix).

Enzymes were separated by horizontal starch gel electrophoresis. All procedures, protocols, and allelic nomenclature follow Murphy et al. (1996b). The analysis utilized 28 enzyme systems encoding 37 presumptive loci. Wherever possible, gene products were resolved on two buffer systems to maximize expression of all variants. Enzyme names, EC numbers, and specific buffer systems for the separation of locus products were the same as those used by Bobyn et al. (1996), Fu et al. (1995), and MacCulloch et al. (1995a). Results obtained from the three Georgian populations were compared to the values obtained from a sample from Gosh, Armenia (Appendix), part of a disjunct population of *L. portschinskii* (MacCulloch et al., 1995a). Alleles from the Gosh sample and from the three Georgian samples were tested for identity in side-by-side comparisons.

Allozyme data were analysed using BIOSYS-1 release 1.7 (Swofford & Selander, 1989). The populations were treated separately for analyses of population variability. All loci were evaluated for three parameters of genetic polymorphism: heterozygosity (MHD), number of alleles per locus (MNA) and percentage of loci which were polymorphic (PLP), and conformity to Hardy-Weinberg expectations using Levene's

(1949) correction for small sample sizes. The populations were examined together for genetic structuring using Wright's (1978)  $F$  statistics. The  $F$  statistics were calculated for all four populations together and for the three Georgian populations alone.

## Results

All four populations were monomorphic for 26 of the 37 loci: mAat-A, Ada-A, Cbp-1, Ck-A, Est-D,  $\beta$ Ga-1,  $\beta$ Ga-2, Gcdh-A, Gda-A,  $\beta$ Glur-A,  $\beta$ Glus-A, Gpi-A, sIdh-A, Ldh-A, Ldh-B, mMdh-A, sMdh-A, sMdhp-A, Mpi-A, Pk-A, Pnp-A, mSod-A, sSod-A, Tpi-A. Genotype frequencies for the 11 polymorphic loci are shown in Table 1. Only one locus (Pep-B) was polymorphic in all four populations. At five of the 11 polymorphic loci, only one population exhibited polymorphism; in most cases this resulted from a single appearance by a rare allele.

At six of the polymorphic loci, at least one of the populations from Georgia exhibited variation, whereas the sample from Gosh was monoallelic. At two loci (mIdh-A and Pgm-A) the Gosh sample exhibited variation whereas all three Georgian samples were monoallelic. Allele frequencies at six loci failed to conform to Hardy-Weinberg probabilities. An excess of heterozygotes was exhibited at Acp-B in Zedazeni and sAcoh-A at Manglisi while mAcoh-A in Manglisi, sAcoh-A, mAcoh-A, and Gpi-B in Kodjori exhibited heterozygote deficiencies.

The indices of genetic variability are shown at the bottom of Table 1. The values of all indices are greater in the three Georgian samples than in the Gosh sample.

The  $F$  statistics for the four populations are as follows:  $F_{IS} = 0.075$ ,  $F_{IT} = 0.365$ ,  $F_{ST} = 0.313$ . Positive values of  $F_{IS}$  and  $F_{IT}$  are indicative of heterozygote deficiencies. The low value of  $F_{IS}$  indicates a slight intrapopulation deficiency. The higher value of  $F_{IT}$  shows an interpopulation heterozygote deficiency greater than the intrapopulation deficiency. The relatively high positive value of  $F_{ST}$  suggests that the four populations do not form a panmictic group. The  $F$ -statistics for the three Georgian populations are  $F_{IS} = 0.079$ ,  $F_{IT} = 0.193$ ,  $F_{ST} = 0.123$ . The first demonstrates a slight intrapopulation heterozygote deficiency virtually identical to that of the four populations. The values of both  $F_{IT}$  and  $F_{ST}$  are much lower than those calculated for the four populations. The lower  $F_{IT}$  shows that removal of the less variable disjunct Gosh

Table 1. Genotype frequencies for polymorphic loci in the four populations of *Lacerta portschinskii*

|                  | Gosh                     | Zedazeni                          | Manglisi                | Kodjori                 |
|------------------|--------------------------|-----------------------------------|-------------------------|-------------------------|
| sAat-A           | aa(39)                   | aa(29)                            | aa(8)                   | aa(5)<br>ab(1)          |
| sAcoh-A          | aa(39)                   | aa(7)<br>ab(13)<br>ac(1)<br>bb(8) | ab(6)<br>bb(2)          | aa(1)<br>bb(3)<br>ac(1) |
| mAcoh-A          | aa(39)                   | aa(17)<br>ab(9)<br>bb(2)          | aa(4)<br>bb(4)          | aa(3)<br>ab(1)<br>bb(2) |
| Acp-B            | aa(39)                   | aa(19)<br>ab(10)                  | aa(7)<br>ab(1)          | aa(3)<br>ab(2)          |
| Cat-A            | aa(38)<br>ab(1)          | ab(3)<br>bb(25)<br>bc(1)          | bb(4)<br>bc(4)          | bb(6)                   |
| Ck-C             | aa(39)                   | aa(26)<br>ab(3)                   | aa(8)                   | aa(6)                   |
| Gpi-B            | aa(39)                   | aa(29)                            | aa(8)                   | aa(5)<br>bb(1)          |
| mIdh-A           | aa(38)<br>ab(1)          | aa(29)                            | aa(8)                   | aa(6)                   |
| Pep-A            | aa(33)<br>ab(6)          | aa(29)                            | aa(2)<br>ab(4)<br>ac(2) | aa(6)                   |
| Pep-B            | aa(31)<br>ab(7)<br>bb(1) | aa(13)<br>ab(7)<br>bb(1)          | aa(3)<br>ac(4)<br>cc(1) | ab(2)<br>bb(2)          |
| Pgm-A            | aa(37)<br>ab(2)          | aa(29)                            | aa(8)                   | aa(6)                   |
| MHD <sup>a</sup> | .011                     | .051                              | .071                    | .039                    |
| MNA <sup>b</sup> | 1.11                     | 1.24                              | 1.19                    | 1.19                    |
| PLP <sup>c</sup> | 5.56                     | 16.22                             | 16.22                   | 16.22                   |

<sup>a</sup>MHD = mean heterozygosity by direct count.

<sup>b</sup>MNA = mean number of alleles per locus.

<sup>c</sup>PLP = percentage of loci polymorphic (0.95 criterion).

population reduces the interpopulational heterozygote deficiency. The lower  $F_{ST}$  indicates that the three Georgian populations are much more likely to constitute a panmictic group than are the four populations. However, this  $F_{ST}$  is still significant ( $p < 0.05$ ), even when corrected for sample size (Workman & Niswander, 1970), demonstrating that the three populations are not panmictic.

## Discussion

### *Disjunct habitats as islands*

Comparison of the three indices of genetic variability shown in Table 1 supports the hypothesis that isolated, disjunct populations exhibit less genetic variation than do larger, contiguous populations. The sample from the disjunct Gosh population has lower values of all three parameters of variability. This demonstrates that terrestrial habitat that is unoccupied, for whatever reason, by a terrestrial species is as effective a barrier to gene flow as is water to insular species.

*Lacerta portschinskii* prefers river valley habitat; the disjunct population is separated by mountains from the southern limit of the species' contiguous range. Gosh is approximately 50 km from the southern limit of the contiguous range and 100 km from the nearest Georgian sampling site. It is not known how long the Gosh population has been isolated; the Caucasus region has undergone many postglacial microhabitat changes, of both natural and human origin, and faunal distributions have changed accordingly. The lower genetic variability could have resulted from a loss of variability in a relict population or from the 'bottle-neck' effect of a founder event.

### *Comparison with other taxa*

Values of MHD, MNA, and PLP of *L. portschinskii* from the contiguous portion of its range approximate those of other Caucasian *Lacerta* sampled from contiguous ranges (*L. caucasica* and *L. daghestanica*, Fu et al., 1995; *L. raddei* and *L. nairensis*, Bobyn et al., 1996; *L. rudis*, MacCulloch et al., 1995a). The disjunct population of *L. portschinskii* has genetic variability parameters equivalent to those found in *L. valentini*, a high-elevation species with patchy distribution (MacCulloch et al., 1995a).

Heterozygosity in *L. portschinskii* from the contiguous part of its range is also comparable to that found in *Podarcis sicula* and *P. melisellensis* from large island and mainland populations (Gorman et al., 1975). Much higher levels of heterozygosity were found in *L. lepida* (Busack, 1987) and in the teiid lizard species *Cnemidophorus tigris* (Gorman, Kim & Taylor, 1977).

A study of 10 genera found mean heterozygosity of 0.01 in fossorial lizards, 0.05 in 'sit-and-wait' species, and 0.09 in vagile species (Gorman, Kim & Taylor, 1977). Heterozygosity in *Lacerta portschin-*

*skii* more closely resembles that of the 'sit-and-wait' group. Similar low heterozygosity values were reported for other bisexual Caucasian *Lacerta* (Bobyn et al., 1996; Fu et al., 1995; MacCulloch et al., 1995a). A more extensive summary reported a mean heterozygosity of 0.051 from 71 lizard taxa or populations, most of which are relatively vagile (Sattler & Ries, 1995). This number is closer to that found in Caucasian *Lacerta*.

In our study, the percentage of loci exhibiting polymorphism was calculated using the 0.95 criterion in BIOSYS-1. Because some studies did not use this criterion, PLP was recalculated without the criterion to facilitate comparison. This produced a different value in the Gosh sample only (PLP = 13.51%); the other populations remained at 16.22%. Mainland populations of *P. sicula* had PLP of 27–45% (Gorman et al., 1975), while *L. lepida* had PLP of 23.1% and 38.5% (Busack, 1987). PLP varied from 10–21% in three species of bisexual *Cnemidophorus* (Dessauer & Cole, 1984).

*Lacerta portschinskii* is one of the parental species of the complex of parthenogenetic *Lacerta*. Among the parthenogenetic species so far examined, clonal diversity is low (Fu et al., in press; MacCulloch et al., 1995b; 1997; Murphy et al., in press). Study of the genetic variability of the parental species is an important part of understanding the origins and diversity of parthenogenetic *Lacerta*.

### Acknowledgements

N. Orlov and A. Agasian assisted in collection of specimens. All collecting and euthanasia were performed under approved animal protocols. Import permits were issued by Agriculture Canada. This study was supported by Natural Sciences and Engineering Council of Canada Grant No. A3148, by the ROM Foundation and the ROM Department of Museum Volunteers (R.W. Murphy), and by the Zoological Institute of the Russian Academy of Sciences, the Russian Scientific and Technical Program 'Priority Trends in Genetics', the International Science Foundation (No. J3Y100), and the Russian Foundation for Basic Science (I.S. Darevsky). This is contribution number 75 from the Centre for Biodiversity and Conservation Biology of the Royal Ontario Museum.

### Appendix

Specimens examined: Georgia, Zedazeni, 41° 50' N, 044° 43' E, ROM 26677–26705 (29 specimens); Georgia, Manglisi, 41° 43' N, 044° 25' E, ROM 26669–26676 (8 specimens); Georgia, Kodjori, 41° 38' 32" N, 044° 41' 02" E, ROM 26663–26668 (6 specimens); Armenia, Gosh, 40° 44' 51" N, 045° 01' 26" E, ROM 23926–23940, 24854–24878 (39 specimens).

### References

- Bobyn, M.L., I.S. Darevsky, L.A. Kupriyanova, R.D. MacCulloch, D.E. Upton, F.D. Danielyan & R.W. Murphy, 1996. Allozyme variation in populations of *Lacerta raddei* and *Lacerta nairensis* (Sauria: Lacertidae) from Armenia. *Amphibia-Reptilia* 17: 233–246.
- Busack, S.D., 1987. Morphological and biochemical differentiation in Spanish and Moroccan populations of the lizard, *Lacerta lepida*. *J. Herpetol.* 21: 277–284.
- Darevsky, I.S., 1967. Rock Lizards of the Caucasus: systematics, ecology and phylogenesis of the polymorphic groups of Caucasian rock lizards of the subgenus *Archaeolacerta*. Nauka, Leningrad. 276 pp. (English translation).
- Darevsky, I.S., L.A. Kupriyanova & T. Uzzell, 1985. Parthenogenesis in reptiles, pp. 411–526 in *Biology of the Reptilia*, vol. 15, edited by C. Gans & F. Billett. Wiley, New York.
- Dessauer, H.C. & C.J. Cole, 1984. Influence of gene dosage on electrophoretic phenotypes of proteins from lizards of the genus *Cnemidophorus*. *Comp. Biochem. Physiol.* 77B: 181–189.
- Fu, J., I.S. Darevsky, R.D. MacCulloch, L.A. Kupriyanova, E.S. Roytberg, T.M. Sokolova & R.W. Murphy, 1995. Genetic and morphological differentiation among Caucasian rock lizards of the *Lacerta caucasica* complex. *Russian J. Herpetol.* 2: 36–43.
- Fu, J., R.D. MacCulloch, R.W. Murphy, I.S. Darevsky, L.A. Kupriyanova & F. Danielyan, in press. The parthenogenetic rock lizard *Lacerta unisexualis*: an example of limited genetic polymorphism. *J. Mol. Evol.*
- Fu, J., R.W. Murphy & I.S. Darevsky, in press. Toward the phylogeny of Caucasian rock lizards: Implications from mitochondrial DNA gene sequences (Reptilia: Lacertidae). *Zool. J. Linn. Soc.*
- Gorman, G.C., M. Soulé, S.Y. Yang & E. Nevo, 1975. Evolutionary genetics of insular Adriatic lizards. *Evolution* 29: 52–71.
- Gorman, G.C., Y.J. Kim & C.E. Taylor, 1977. Genetic variation in irradiated and control populations of *Cnemidophorus tigris* (Sauria, Teiidae) from Mercury, Nevada, with a discussion of genetic variability in lizards. *Theor. Appl. Genet.* 49: 9–14.
- Levene, H., 1949. On a matching problem arising in genetics. *Ann. Math. Stat.* 20: 91–94.
- MacCulloch, R.D., J. Fu, I.S. Darevsky, F. Danielyan & R.W. Murphy, 1995a. Allozyme variation in three closely related species of Caucasian rock lizards (*Lacerta*). *Amphibia-Reptilia* 16: 331–340.
- MacCulloch, R.D., R.W. Murphy, L.A. Kupriyanova & I.S. Darevsky, 1995b. Clonal variation in the parthenogenetic rock lizard *Lacerta armeniaca*. *Genome* 38: 1057–1060.
- MacCulloch, R.D., R.W. Murphy, L.A. Kupriyanova & I.S. Darevsky, 1997. The Caucasian rock lizard *Lacerta rostombekovi*: a monoclonal parthenogenetic vertebrate. *Biochem. Syst. Ecol.* 25: 33–37.

- Murphy, R.W., I.S. Darevsky, R.D. MacCulloch, J. Fu & L.A. Kupriyanova, 1996a. Evolution of the bisexual species of Caucasian rock lizards: a phylogenetic evaluation of allozyme data. *Russian J. Herpetol.* 3(1): 18–31.
- Murphy, R.W., I.S. Darevsky, R.D. MacCulloch, J. Fu, L.A. Kupriyanova, D.E. Upton & F. Danielyan, in press. Old age, multiple formations or genetic plasticity? Clonal diversity in a parthenogenetic Caucasian rock lizard, *Lacerta dahli*. *Genetica*.
- Murphy, R.W., J.W. Sites, Jr., D.G. Buth & C.H. Haufler, 1996b. Proteins: Isozyme electrophoresis, pp. 51–120 in *Molecular Systematics* (2nd edition), edited by D.M. Hillis, C. Moritz & B.K. Mable. Sinauer Associates, Sunderland, Mass.
- Sattler, P.W. & J.S. Ries, 1995. Intraspecific genetic variation among four populations of the Texas horned lizard, *Phrynosoma cornutum*. *J. Herpetol.* 29: 137–141.
- Swofford, D.L. & R.B. Selander, 1989. BIOSYS-1: A computer program for the analysis of allelic variation in population genetics and biochemical systematics, Release 1.7. Illinois Nat. Hist. Survey, Urbana.
- Workman, P.L. & J.D. Niswander, 1970. Population studies on southwestern Indian tribes. II. Local genetic variation in the Papago. *Am. J. Human Genet.* 22: 24–49.
- Wright, S., 1978. *Evolution and genetics of populations*, vol. 4. Variability within and among natural populations. Univ. Chicago Press, Chicago.